Down Syndrome Screening Sample Details:

		First Trimester Combined Down Syndrome Screening	Second Trimester Biochemical Down Syndrome Screening
Window for Sample Collection		11w0d to 13w6d	15w0d to 20w0d
Primary Sample	**Type of Sample**	Venous blood	
	Type of Container	Clotted blood tube (gel separator preferred) Vacuette gel separation clotted blood tube (Z Serum Sep Clot Activator) is recommended. Fig.1	
	Sample Volume	4 mL or above	
	Storage Temperature after collection (if delivery within 2 hours after collection is not possible)	2-8°C	
	Sent to Laboratory after collection	Within 24 hours	
	Identification	At least 2 unique identifiers clearly on tube, leaving a window of 0.5cm gap on tube for viewing of blood level. Fig.2	
Secondary Sample	**Type of Sample**	Serum	
	Sample Volume	2mL or above	
	Storage after serum separation	2-8°C	
	Sent to Laboratory after collection	Within 72 hours	
	Identification	At least 2 unique identifiers clearly on tube, leaving a window of 0.5cm gap on tube for viewing. Fig.2	

Fig.1 Blood sample collected in Vacuette gel separation clotted blood (Z Serum Sep Clot Activator).

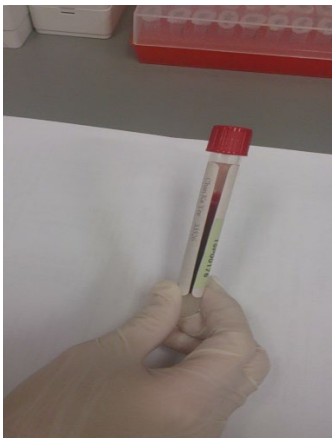

Fig.2 Affix patient's label on the tube leaving a 0.5cm gap for viewing the blood level.

Sample Transportation:

	First Trimester Combined Down Syndrome Screening	**Second Trimester Biochemical Down Syndrome Screening**
1.	Check the cap of the primary tube close tightly and seal the secondary tube with parafilm to avoid spillage and soiling.	
2.	Check the Laboratory Request Form completely filled.	
3.	Put the Laboratory Request Form with the sample in a specimen carrier bag. Fig. 3	
4.	• Blood sample transport in room temperature if it arrives the Laboratory within 2 hours after blood drawn. • Transport the sample on ice/coolant gel pad in a cooler bag to the Laboratory from 2 to 24 hours after blood drawn. Fig. 4 (For HA centres, please include a Sample Delivery Checklist with patient labels for acknowledgement of samples on arrival.)	
5.	Send to laboratory before 15:00 daily when open.	

Fig.3 Place blood sample and the form in a specimen carrier bag.

Fig.4 Transport on ice/coolant gel pad in a cooler bag

Handling Procedure of Biological Spill

- All specimens are considered as biohazardous. It is generally agreed that a waiting time of 30 minutes should be observed before attempting to clean up a complex spill. This allows large particles to settle and smaller ones to be removed by ventilation system. It is therefore recommended to wait at least 30 minutes before cleaning up is commenced.
- Alert people in immediate area of spill.
- Put on laboratory gown, disposable gowns, N95/100 mask, full-face shield or eye shield, disposable cap, latex gloves before doing the decontamination procedure.
- Cover spill with paper towels, Haz-Mat PIG Mat or other absorbent materials.
- Carefully pour 10000 ppm sodium hypochlorite around the edges of the spill and then into the spill. Avoid splashing.
- Allow a 10 minutes contact time.
- Use paper towels to wipe up the spill, working from the edges into the center.
- Clean up spill area with fresh paper towels soaked in disinfectant.
- Discarded all the used absorbent materials or paper towels into biohazard bag.
- Soiled Laboratory Request Form should not be exposed but be kept in the specimen bag. Seek sender to fill in another Request Form and send by fax.
- Severe leakage sample should not be accepted for processing.
- Report the incidence to Quality Manager or deputy.